Progesterone and progesterone receptors (PRs) are essential for the development and cyclical regulation of hormone-responsive tissues including the breast and reproductive tract. Altered functions of PR isoforms contribute to the pathogenesis of tumors that arise in these tissues. In the breast, progesterone acts in concert with estrogen to promote proliferative and pro-survival gene programs. In sharp contrast, progesterone inhibits estrogen-driven growth in the uterus and protects the ovary from neoplastic transformation.

Progesterone-dependent actions and associated biology in diverse tissues and tumors are mediated by two PR isoforms, PR-A and PR-B. These isoforms are subject to altered transcriptional activity or expression levels, differential crosstalk with growth factor signaling pathways, and distinct post-translational modifications and cofactor-binding partners. Herein, we summarize and discuss the recent literature focused on progesterone and PR isoform-specific actions in breast, uterine, and ovarian cancers. Understanding the complexity of context-dependent PR actions in these tissues is critical to developing new models that will allow us to advance our knowledge base with the goal of revealing novel and efficacious therapeutic regimens for these hormone-responsive diseases.

**Key Words**
- progesterone
- progestin
- progesterone receptor
- isoforms
- breast cancer
- endometrial cancer
- uterine
- ovarian cancer

**Introduction**

Progesterone and progesterone receptors (PRs) are increasingly gaining attention for their emerging role as critical regulators of breast and gynecological cancers. With this newfound spotlight on PR action, there is an urgent need to define the mechanisms by which progesterone and progestins exert their effects upon tumor types in different PR+ tissues and bring clarity to areas of confusion in the field. Much of the difficulty lies in the nuanced context-dependent actions of PR, the different isoform-specific actions of PR-A relative to PR-B (PR isoforms are not measured separately in the clinic), the differential or potential off-target actions of synthetic progestins (i.e., used clinically) vs natural progesterone, and the seemingly paradoxical biological effects that progesterone exerts on the breast vs gynecological tissues. In the breast, progesterone is proliferative and works in concert with estrogens and estrogen receptors (ERs) to induce expansion of glandular structures during development (reviewed in Brisken & O’Malley (2010)). Progesterone is a key mediator of mammary gland stem cell expansion (Brisken 2013). In the presence of estrogen, ER-induced expression of PR is required to induce proliferation by both autocrine and paracrine mechanisms (Brisken 2013); PR-target genes include secreted factors (wnt4) that act on nearby...
PR-negative cells. In contrast to ER and PR cooperative actions in the breast, progesterone opposes ER actions in the ovary and endometrium, and acts in an antiproiferative manner to induce tissue regression (Kim & Chapman-Davis 2010). Our goal herein is to examine the reproductive tract and breast tissues (Lydon 1995, Graham & Clarke 1997). Pregnancy interrupts this process and is characterized by high progesterone levels, which are required for fetal development, breast development for lactation, maintenance of uterine/placental integrity, and myometrial quiescence (Mendelson 2009).

Expression of PR in responsive tissues is driven by estrogen-bound ER, and therefore, ER is permissive for the actions of PR and progesterone. As a result, one experimental challenge that researchers face in determining the actions of PR is their differentiation from those of ER. Elegantly designed mouse models and transplant studies have delineated the developmental processes attributed to each receptor (reviewed in Brisken & O’Malley (2010)). Briefly, PR-B is the predominant isoform required for mammary gland development and expansion, whereas PR-A is necessary for proper uterine development and reproductive actions (Conneely et al. 2001). PR expression is limited to 10–15% of mammary luminal cells and primarily signals in a paracrine manner to induce proliferation of steroid receptor-negative cells (Brisken et al. 1998). PR-containing cells proliferate autonomously during periods of massive glandular expansion, such as pregnancy. PR is expressed in both the epithelial and stromal compartments of the breast and uterus and signals in both paracrine and autocrine manners to affect biology (Kim & Chapman-Davis 2010, Brisken 2013, Kim et al. 2013). The actions of progesterone and its isoforms in normal physiology of the breast (see Kariagina et al. (2008)), uterus (see Kim et al. (2013)), and ovary (see Modugno et al. (2012)) have been extensively reviewed previously.

PRs are members of the steroid hormone family of nuclear receptors and as such are composed of a modular domain structure that includes an N-terminal domain (NTD), DNA-binding domain (DBD), hinge region (H), and hormone-binding domain (HBD) (Fig. 1). There are three PR isoforms: full-length PR-B, N-terminally truncated PR-A (−164 amino acids), and the non-functional PR-C, consisting of only the hinge region and HBD (Fig. 1). PR-B and PR-A are typically expressed in equimolar ratios and function as ligand-activated transcription factors, whereas expression of PR-C is limited and may serve largely to sequester ligand, as it is incapable of binding DNA (Condon et al. 2006).

Progesterone diffuses through the lipid membrane and interacts with the HBD of PR-A or PR-B. This interaction alters the confirmation of PR favoring nuclear localization, dimerization (A:A, B:B, or A:B dimers are possible), and DNA binding. Classically, PR binds progesterone response elements (PREs) in the DNA and recruits cofactors and transcriptional machinery to initiate gene transcription. PR can also participate in the transcriptional complexes of other DNA-bound transcription factors to alter gene expression (Owen et al. 1998, Stoecklin et al. 1999, Cicatiello et al. 2004, Faivre et al. 2008). Non-classical or extranuclear signaling of PR involves direct binding of PR to kinases complexed at the membrane with growth factor receptors (such as EGFR or IGF1R) to initiate rapid activation of downstream signaling cascades (Migliaccio et al. 1998, Boonyaratankornkit et al. 2001). For example, progesterone induces rapid activation of ERK1/2 MAPK pathways, which function to activate a variety of transcription factors via phosphorylation events, including PR itself (Migliaccio et al. 1998, Boonyaratankornkit et al. 2001, Faivre et al. 2008; Fig. 1). Notably, PR-B, but not PR-A, is capable of rapid signaling, probably in part owing to its relatively increased occupancy in the cytoplasm (Boonyaratankornkit et al. 2007). The regulation of gene programs driven by PR and progesterone is highly dependent on the local cellular environment and the intracellular signaling context. Thus, PRs act as ‘sensors’ of cell context, rapidly adjusting to hormonal fluctuation and integrating a variety of extracellular signals to enable tight control of developmental programs. The mechanisms by which PR selects and modulates genetic programs in response to variable external signals are discussed later in this review.

Breast

Proliferative actions of PR in the breast

Progesterone, acting through PR, is a critical mediator of mammary gland tissue expansion during breast

http://jme.endocrinology-journals.org
DOI: 10.1530/JME-14-0252
© 2015 Society for Endocrinology
Published by Bioscientifica Ltd.
development after puberty. Mouse models lacking PR-B, but not PR-A, exhibit marked defects in mammary gland branching and alveologenesis (Conneely et al. 2003), supporting the concept that PR-B is the predominant isoform required for mammary gland development and expansion. Interestingly, ER is also required for mammary gland proliferation during pubertal development (Daniel et al. 1987). However, in the adult mammary gland, proliferation occurs via the actions of PR at its primary target genes, while ER is necessary for PR expression (Brisken 2013). In hormone ablation and replacement studies in adult mice, ovariectomy arrests glandular proliferation. Exogenous estrogen alone provides a weak signal, whereas treatment with estrogen and progesterone restores glandular proliferation (Wang et al. 1990). Tissue transplant studies in genetically modified mice demonstrated that ER and PR induce proliferation in mammary gland structures primarily via paracrine signaling (Brisken et al. 1998). PR is expressed in both the epithelial and stromal compartments of the breast and is limited to 10–15% of mammary luminal cells (Brisken 2013). Progestin stimulation of PR-positive mammary epithelial cells induces transcription and secretion of multiple mitogenic factors, including Wnts, Areg, HB-EGF, and receptor activator of nuclear factor kappaB ligand (RANKL) that induce proliferation of neighboring PR-negative cells (Brisken 2013; Fig. 2).

The color of phosphorylation sites is associated with the following: red, MAPK; green, CDK2; yellow, CK2; purple, unknown kinases. PR, PR protein isoforms A, B, or C; NTD, N-(amino)-terminal domain; DBD, DNA-binding domain; H, hinge region; HBD, hormone-binding domain; AF, activation function 1–3; P, phosphorylation; A, acetylation; SUMO, small ubiquitin-like modifier (SUMOylation). Dysregulation of Kinase Inputs to PR in Cancer: the percent of The Cancer Genome Atlas Research Network 2011 (TCGA) tumors containing alterations in MAPK, CDK2, or CK2 components was identified using the cBioPortal.org analysis tool. For analysis of dysregulated kinases: MAPK includes canonical c-Raf, Mek, and Erk signaling pathway genes: RAF1, MAP3K1, MAP3K2, MAPK3, MAPK1; CDK2, cyclin-dependent kinase 2; CK2, casein kinase 2; CSNK2A1, casein kinase 2, α1 polypeptide.

Figure 1

The post-translational modifications of progesterone receptors. Seventeen post-translational modification sites that affect PR-mediated transcriptional action. PR-B, but not PR-A, includes 164 additional amino acids in the NTD (called B upstream segment), where the third activation function domain and multiple phosphorylation sites are located. PR-B and PR-A are transcribed from the same gene and their protein isoforms are identical to amino acids 165–993. The protein tertiary structure results in a folding at the hinge region between the DBD and HBD. Post-translational modifications (phosphorylation, acetylation, and SUMOylation) can occur basally or in response to ligand binding and affect PR transcriptional activity. In particular, activated protein kinase pathway input to PR via phosphorylation and these pathways are heavily altered in breast, ovarian, and uterine carcinomas. Numbering reflects amino acid residue positions.

The post-translational modifications of progesterone receptors. Seventeen post-translational modification sites that affect PR-mediated transcriptional action. PR-B, but not PR-A, includes 164 additional amino acids in the NTD (called B upstream segment), where the third activation function domain and multiple phosphorylation sites are located. PR-B and PR-A are transcribed from the same gene and their protein isoforms are identical to amino acids 165–993. The protein tertiary structure results in a folding at the hinge region between the DBD and HBD. Post-translational modifications (phosphorylation, acetylation, and SUMOylation) can occur basally or in response to ligand binding and affect PR transcriptional activity. In particular, activated protein kinase pathway input to PR via phosphorylation and these pathways are heavily altered in breast, ovarian, and uterine carcinomas. Numbering reflects amino acid residue positions.
Recent findings have implicated progesterone as a key mediator of breast cancer progenitor cell plasticity. Exposure of ER+/PR+ breast cancer cultures to progesterone induces the emergence of cells expressing known progenitor and stem cell markers, such as CK5 (KRT5) and CD44. These cells possess properties that include therapy resistance and heightened mammosphere-forming potential (Horwitz et al. 2008, Cittelly et al. 2013). PR regulation of microRNAs (miRs) facilitates the increase in stem-like phenotypes in breast cancer cell cultures. Downregulation of miR-29a facilitates dedifferentiation by releasing the transcription factor KLF4 to alter gene programs (Cittelly et al. 2013), while miR-141 repression by PR prevents downregulation of PR itself and STAT5 (STAT5A), which is known to control progenitor cell phenotypes (Yamaji et al. 2009). Notably, evidence is mounting in favor of the prevailing hypothesis that hormone replacement therapies (HRTs), which include progestins, induce a greater incidence in breast cancer by the expansion of pre-malignant stem cell populations (Horwitz & Sartorius 2008).

Indeed, epidemiological evidence and clinical findings have demonstrated that synthetic progestins, whether given in HRT as post-menopausal treatments or as hormonal contraceptives in pre-menopausal women, confer a greater breast cancer risk. Progesterin-containing contraception is linked to an increased risk of developing breast cancer in multiple epidemiological studies (Collaborative Group on Hormonal Factors in Breast Cancer 1996, Hunter et al. 2010, Li et al. 2012, Soini et al. 2014). Similarly, other epidemiological studies indicate that greater exposure to progesterone throughout an individual’s lifetime leads to greater likelihood of breast cancer (reviewed in Knutson & Lange (2014)). Large-scale clinical trials, including the Women’s Health Initiative (hazard ratio 1.26; 95% CI 1.02–1.55) (Chlebowski et al. 2009), Million Women’s Study (relative risk 2.00 (1.88–2.12), P < 0.0001) (Beral 2003), E3N-EPIC cohort (relative risk 1.3 (1.1–1.5)) (Fournier et al. 2005), and Finnish Cancer Registry case-controlled analysis (odds ratio 1.36; 95% CI 1.27–1.46) (Lyytinen et al. 2010), demonstrate that women taking progestins added to estrogen therapy are at greater risk of developing breast tumors. Unexpectedly, estrogen alone as a single-agent therapy for women who have had a hysterectomy confers protection against breast cancer (hazard ratio 0.77; 95% CI 0.62–0.95) (Chlebowski & Anderson 2012). Recently, a retrospective analysis of Finnish women using the levonorgestrel-releasing intrauterine system of contraception has also demonstrated an increased risk of breast cancer (standardized incidence ratio for one purchase 1.16; 95% CI 1.09–1.22. For users with two purchases: 1.40; 95% CI 1.24–1.57) (Soini et al. 2014). However, the same regimen conferred protection from endometrial (for one purchase 0.50; 95% CI 0.35–0.70; for users with two purchases 0.25; 95% CI 0.05–0.73) and ovarian cancers (0.60; 95% CI 0.45–0.76) as well as lung (0.68; 95% CI 0.49–0.91) and pancreatic (0.50;
95% CI 0.28–0.81) cancers (Soini et al. 2014). In studies comparing HRT containing synthetic and natural progestins (albeit with smaller cohort sizes), natural progestins did not significantly increase breast cancer risk (Fournier et al. 2005, 2008). Importantly, the relative instability of natural progesterone may account for the differential biological outcomes compared with long-lived synthetic progestins, raising interesting questions about the duration and level of exposure to PR activators (reviewed in Brisken (2013)). Alternatively, synthetic progestins may elicit off-target effects on other steroid receptors that may also contribute to their deleterious or protective effects (reviewed in Knutson & Lange (2014)). Together, these epidemiological and clinical findings support the notion that uncontrolled PR action in pre-neoplastic breast tissue contributes to breast cancer development. These data are corroborated by an expansive body of literature demonstrating in both in vivo and in vitro models of luminal breast cancer that exposure to progestins increases proliferation and promotes pro-survival and progression of malignant breast cells (reviewed in Daniel et al. (2011)). Interestingly, while ~70% of newly diagnosed breast tumors are ER+/PR+ (luminal-type tumors), ~40 and 25% of luminal tumors exhibit loss of heterozygosity (LOH) at the PGR or ER locus respectively (Knutson & Lange 2014). Generally, ER and PR LOH are positively correlated. However, interestingly, despite this genetic loss, ER and PR mRNA levels remain very similar to that of diploid luminal tumors (Knutson & Lange 2014), suggesting that other compensatory factors may exist in these tumors to maintain ER and PR expression.

**Context-dependent PR activation**

The gene programs driven by PR are determined by a diverse array of cellular conditions that modify the receptor and its cofactors, which serve to direct transcriptional complexes to specific promoters. Not surprisingly, progesterone binding produces a dramatic shift in PR-mediated gene selection. PR remains bound to and regulates expression (both activation and repression) of a multitude of genes in the unliganded state (Knutson et al. 2012a, Daniel et al. 2014, Dressing et al. 2014), whereas PR relocates to a subset of progesterone-responsive genes upon ligand binding. These two broad categories of PR-driven genes, unliganded and liganded gene sets, are further regulated by the convergence of particular kinase pathway outputs (Fig. 1), in the form of direct phosphorylation of PR and its cofactors (reviewed in Hagan & Lange (2014)). For example, phospho-S294 PR, in response to MAPK or CDK2 activation, regulates an overlapping yet distinct set of gene targets in the presence of progesterone compared with phospho-S81 PR (via activated CK2), and the same (i.e., sensitivity of selected genes to phosphorylated PR) is true for unliganded target genes (Daniel et al. 2007, Daniel & Lange 2009, Hagan et al. 2011a, Knutson et al. 2012b). To date, post-translational modifications identified on PR that alter its transcriptional activity include: phosphorylation (S294, S345, S81, and S400), SUMOylation (K388), acetylation (K183, K638, K640, and K641), and ubiquitylation (Fig. 1; Lange et al. 2000, Pierson-Mullany & Lange 2004, Daniel et al. 2007, 2010, Faiivre et al. 2008, Daniel & Lange 2009, Beleut et al. 2010, Hagan et al. 2011a, Knutson et al. 2012b, Chung et al. 2014, Dressing et al. 2014). PR transcriptional activity and promoter selection are thus dramatically altered by the activation state of mitogenic signaling pathways such as MAPK, AKT, CDK2, cAMP, and CK2 (Fig. 1). In addition, the availability of particular cofactors and their post-translational modification states are also determinants of PR gene selectivity (Hagan & Lange 2014). In short, PR is capable of inducing diverse biological outcomes dependent on the cellular context as determined by the presence or absence of activated signaling pathways and the availability of cofactors. Studies probing the complexity of PR action thus require particular care in both the design of model systems and the interpretation of specific results. For example, breast cancer cells in culture respond differently to progestins depending on the culture conditions. Cells cultured in 2D (adherent to plastic dishes) elicit a biphasic response characterized by one or few rounds of cell cycle progression followed by growth arrest (Musgrove et al. 1991, Groshong et al. 1997), whereas in 3D culture conditions (such as soft agar) progesterone is clearly mitogenic and a mediator of cell survival (Faiivre & Lange 2007). These data may reflect an alteration in signaling pathways and kinase activation that is dependent upon cell polarity and/or cellular junctions or ‘structural’ communication that in turn informs PR gene selectivity and modulates the strength and duration of its transcriptional activity (i.e., aspects of PR action that are missed using reporter assays).

Notably, PR-A and PR-B are differentially susceptible to post-translational modifications in response to the same kinase signals. This complexity contributes to the distinctions between the genes they activate and ultimately the biological consequences for PR+ and nearby PR-null cells (i.e., responsive to PR-derived paracrine signals). For example, PR-B, but not PR-A, is robustly phosphorylated on Ser294 in response to MAPK activation.
Ser294 phosphorylation is a major regulatory input for PR-B, controlling increased sensitivity to progestin, an increased rate of ubiquitylation of PR (an activation step for several steroid receptors (Salghetti et al. 2001)) required for degradation through the 26S proteasome pathway (Lange et al. 2000), decreased SUMOylation on K388 (Daniel et al. 2007), unliganded transcriptional activity (Daniel & Lange 2009), and altered promoter selectivity (Knutson et al. 2012a). Similarly, CUE domain containing 2 (CUEDC2), an ubiquitin-binding motif-containing protein, targets the K388 SUMOylation site for ubiquitylation and degradation of PR, suggesting that PR ubiquitylation may oppose SUMOylation via competition for the same required lysine residue (Zhang et al. 2007). In contrast, modest (low to unmeasurable in intact cells) PR-A Ser294 phosphorylation confers less responsiveness of this isoform to kinase inputs and increased K388 SUMOylation (a transcriptionally repressive modification) (Daniel et al. 2007). The increased SUMOylation of PR-A relative to PR-B may account for the increased trans-repressive activity of this isoform (Abdel-Hafiz et al. 2009). PR-A is known to repress the activities of PR-B, ER, androgen receptor (AR), and gonadotropin receptor (GR) (Abdel-Hafiz et al. 2002). PR isoforms also participate in distinct complexes with cofactors, owing in part to differences in post-translational modifications, and also due to cofactor-binding sites located in the PR-B N-terminus (Giangrande et al. 2000). Differential transcriptional complex components aid in determining relative transcriptional activities (i.e., altered hormone sensitivity) and are responsible for directing receptor gene selectivity; PR-A and PR-B have distinct and overlapping gene signatures in breast cancer cells (Richer et al. 2002).

Importantly, evaluation of endogenous genes to determine the impact of phosphorylation events on steroid receptor action is critical. Phosphorylation events have been shown to alter promoter selection rather than absolute transcriptional activity. Luciferase assays measure transcriptional activity, but fail to detect alterations in promoter selectivity. Thus, mutant PRs that appear to be fully functional in luciferase assays repeatedly fail to activate selected endogenous (native) promoters of genes in intact cells (Qiu & Lange 2003, Daniel et al. 2009).

In breast cancer cell models and clinical studies, the ratio of PR-A to PR-B is a critical determinant of the biological or physiological response to progesterone (reviewed in Mote et al. (2007)). In normal tissues, PR-A and PR-B typically occur as a 1:1 ratio. However, unbalanced PR-A and PR-B expression occurs in the normal breast of women at high risk of developing breast cancer, while altered ratios in breast tumors are linked to endocrine resistance (Venkitaraman 2002, Mote et al. 2004). Differential signaling and transcriptional activities of the isoforms as well as altered ability of PR-A to transrepress other steroid receptors probably contribute to breast pathologies (Abdel-Hafiz et al. 2002). The mechanisms that drive imbalanced PR-A-to-PR-B ratios are still under investigation. We hypothesize that increased kinase activity in the pre-malignant or early malignant setting drives PR-B phosphorylation leading to its hyperactivity and the subsequent rapid protein turnover (relative to PR-A) (Lange et al. 2000, Daniel et al. 2007). Thus, activated phospho-PR-B receptors exhibit an overall decreased steady-state protein level relative to PR-A receptors (which are not appreciably phosphorylated on Ser294 in response to MAPK or CDK2 activation). In this setting, PR-B exhibits heightened transcriptional activity on selected target genes, yet is less detectable. PR-B is widely recognized as the more proliferative isoform (Faiivre & Lange 2007) and, as such, may primarily drive the dysplastic phenotypes observed in these tumors. In addition, loss of PR-A (the more repressive isoform) via promoter methylation (Pathiraja et al. 2011) may lead to loss of its protective actions and provide an epigenetic ‘stepping stone’ in tumor progression, an event that is similarly observed in endometrial tumors (discussed later in this review). Unfortunately, in the clinic, total PR levels are still measured using antibodies that fail to distinguish between PR isoforms (primarily conducted by immunohistochemistry (IHC)). This represents a missed opportunity to gain a much better understanding of PR isoforms as distinct biomarkers of disease progression. Given the differential activities of the receptors and their known effects on breast cancer cell biology, measuring the isoforms individually is likely to provide valuable information relevant to the use of tailored endocrine therapies. In addition, examining PR isoform-specific gene programs in tumors may further inform tumor biology and in turn drive treatment strategies targeting individual PR isoforms.

**ER and PR crosstalk**

An emerging paradigm in steroid receptor biochemistry is crosstalk between different receptor types, which allows receptors to modulate the signaling and transcriptional responses to non-cognate ligands. Recent studies have demonstrated that steroid receptors, including PR, ER, AR, and GR, participate in complexes with each other to a degree that is much more extensive than considered previously (Peters et al. 2009, Giulianelli et al. 2012,
Need et al. 2012, Daniel et al. 2014). This crosstalk is critical to understanding breast cancer biology because ER and PR are capable of modulating the activities of each other, which has implications for endocrine therapy responses. Our recent studies have demonstrated that ER, PR-B, and the coactivator and signaling scaffold molecule, PELP1, are constitutively complexed in human breast tumor samples and cell lines (Daniel et al. 2014). The consequences of this interaction in the presence of estrogen in ER+/PR+ breast cancer cell models include: enhanced ER phosphorylation, altered ER promoter selectivity, increased cellular proliferation, and decreased sensitivity to tamoxifen treatment (Daniel et al. 2014). In similar studies, ER and PR complexes exhibited enhanced transcriptional and proliferative responses to progestins as well (Giulianelli et al. 2012). Ultimately, these studies demonstrated that breast cancer cells harboring both ER and PR-B might, in fact, be exquisitely sensitive to exposure of either hormone. Perhaps, in the case of endocrine resistance, steroid receptors can substitute for each other or utilize alternative ligands to drive proliferative gene programs and escape inhibition of one receptor type. Relevant to this concept, ER-α may be activated by thyroid hormone (T₄) or by cholesterol metabolites (Tang et al. 2004, Wu et al. 2013), providing an easy ‘escape’ for tumors under the selection pressure of aromatase inhibitors.

PRs ‘enable’ signaling pathways via ‘feed-forward’ cofactor expression

Our recent studies have elucidated mechanisms by which PR acts as a sensor to integrate multiple signals (kinase pathway activation and hormone exposure) and ensure persistent activation of particular gene programs, in part via regulation of unique cofactor expression and by upregulation of signaling pathway components. For example, STAT5 is a PR target gene (Richer et al. 1998), and these factors interact directly and cooperate at numerous PR/STAT5 target genes (Hagan et al. 2013). Progesterone binding in the presence of high intracellular CK2 activity, a commonly activated kinase in cancer, initiates PR phosphorylation on Ser81 to induce robust STAT5 expression. PR then cooperates with STAT5 on selected target genes required for proliferation, stem cell maintenance, and inflammatory responses (Hagan et al. 2011a). In fact, we hypothesized that STAT5 functions as a pioneer factor recruiting S81 phosphorylated PR to specific chromatin loci (Hagan & Lange 2014). In other circumstances, namely during cell cycle progression through mitosis when both MAPK and CDK2 phosphorylation sites on PR are induced (Ser294, Ser345, and Ser400), cyclin D1 mRNA and protein are directly upregulated in response to progestin (Dressing et al. 2014). Phospho-S345 PR and cyclin D1 (acting as a coactivator of transcription) then cooperate as part of SP1-containing transcriptional complexes to enact a new genetic program in the cell, distinct from that of cells with little to no cyclin D1 expression (Dressing et al. 2014). This paradigm, whereby PR induces the same pathway factors that are required to fulfill specific context-dependent biological outcomes in response to progestin, is recapitulated in ovarian cells. In progesterone-treated ovarian cancer cell models, PR induces the increased expression of FOXO1, which in turn binds to PR in order to further modulate selected FOXO1/PR target genes required for progestosterone-dependent induction of cellular senescence (Diep et al. 2013) (discussed later in this review). In addition, PRs are exquisitely sensitive to the local signaling environment in addition to ligand availability and the presence of cofactors that, when bound to PR, persistently direct or select highly specific genetic programs. The potential for distinct biological responses to transient vs persistent exposure to progestins is not considered clinically, for example, during HRT. The kinetics of feed-forward signaling events enacted by ligand-bound PRs is unknown and a topic for further study.

Uterus

Epidemiological role of progesterone in endometrial cancers

Continuous exposure to sex steroid imbalances, where there is insufficient progesterone or excessive estrogen acting upon endometrial tissue, can result in hyperplasia of the glandular epithelial tissue, with the potential to progress to atypical hyperplasia and endometrial carcinoma (Yang et al. 2011, Kim et al. 2013). Endometrial cancer is the most common gynecologic cancer and is classified into type I and type II carcinomas, each characterized by varied hormonal dependence, glandular/stromal architecture, progression, and patient outcome (Samarnthai et al. 2010). Type I endometrioid tumors represent 70–80% of all endometrial cancers, often estrogen dependent, presenting at a lower grade at an early stage with good patient prognosis. Type II non-endometrioid tumors are aggressive and rarely hormone dependent, diagnosed at a later stage with poorer prognosis and higher recurrence rates. In the progression
from low grade (well-differentiated cancers with clear glandular structures and stromal tissue) to high grade (poorly differentiated cancers), loss of stromal tissue and myometrial invasion is common. Owing to the ability of progesterone to antagonize proliferation and promote atrophy of the endometrium (Charles 1964), progesterone and its derivatives (progestins) have been used successfully as therapeutics to treat endometrial hyperplasias and cancers. High response rates (70–90%) are often observed for women with pre-invasive atypical hyperplasia or early stages of endometrial cancers without myometrial invasion (Kaku et al. 2001, Ushijima et al. 2007). Yet, the efficacy of progestins declines to modest response rates (15–25%) when used for cases of advanced or recurrent cancer (Banno et al. 2012) and more than 30% of patients with well-differentiated, hormone-dependent type I tumors will fail to respond (Shao 2013). The mechanisms that result in the progression from progestin sensitivity to the hormone refractory state, or ‘progesterone resistance’, are poorly understood.

Unlike most mammals, the uterine endometrium of human and some non-human primates undergoes cyclical monthly changes that result in the growth, angiogenesis, and differentiation of the functional (proliferative) endometrium (Ramsey et al. 1976, Clancy 2009). Shifts in the synthesis and secretion of the ovarian steroids, estrogen and progesterone, during this menstrual cycle serve as the principal hormonal drivers for these changes. Rising circulating estradiol during the mid- to late follicular phase of the cycle promotes the proliferation of the functional endometrium (Fig. 3); this most luminal portion of the endometrium regenerates each cycle from the basal endometrium and contains the glandular epithelial and stromal cells. Following ovulation, during the secretory luteal phase, rising circulating progesterone antagonizes these proliferative effects of estradiol and

**Figure 3**

Epithelial–stromal interactions regulating proliferation and differentiation of the uterine endometrium. The uterine endometrium is stylized in this figure, with predominant signaling pathways represented during the proliferative follicular phase of the menstrual cycle (above dotted line) and during the differentiation of the luteal phase (below dotted line). Arrows on the right indicate relative concentrations of circulating steroid hormone levels. During the follicular phase, the predominant steroid, estrogen (E2; estradiol), acts through its receptor (ER; expressed in epithelium and stroma) to activate the PI3K/Akt pathway and promote inhibitory phosphorylation of GSK-3β, leading to activation of Wnt signaling, regulation of cell cycle proteins, and enhanced cell proliferation. E2 can also induce the expression of critical growth factors such as Wnt ligands, IGF1, and FGFs that are secreted by the epithelia and stroma, and which bind to epithelial membrane receptors (i.e., receptor tyrosine kinases, RTKs) to support proliferation. During the luteal phase and early pregnancy, progesterone (P4), as the predominant hormone, antagonizes E2-induced proliferation and promotes differentiation of the glandular epithelium. P4 acts through its receptor (PR) to induce expression of Indian hedgehog (IHH) within the epithelium, which binds to patched (PTCH) on the surface of the stromal cells and through the COUP-TFII and Hand2 complex inhibits expression of FGFs. In addition, P4 also appears to induce the stromal expression of the Wnt signaling antagonist, dickkopf-related protein 1 (DKK1) and the transcription factor, FOXO1, which leads to inhibition of Wnt signaling, inhibition of cell cycle progression, and expression of decidualization-specific genes for stromal cell differentiation. Frequent alterations in endometrial cancer include altered ER/PR expression, PTEN loss of function, activation of PI3K/AKT signaling, and mutations to FGFR; these events are predicted to affect PR actions in the context of tumorigenesis.
supports the differentiation of stromal cells and the decidualization of the endometrium (Fig. 3).

Epithelial–stromal interactions within the endometrium: PR isoform specificity

PR-A and PR-B are expressed in both the epithelial and the stromal cells of the endometrium and their expression fluctuates during the menstrual cycle as well as during implantation and pregnancy. During the follicular phase of the cycle, both isoforms are expressed at high levels when the endometrium is proliferating, then decline after ovulation through the luteal phase (Mylonas et al. 2007). In general, PR-A expression appears to be predominant in the stromal cells, declining less during the luteal phase whereas, in glandular epithelial cells, PR-B dominance is observed in the late secretory phase (Mote et al. 1999). The antagonistic effects of progesterone on the estrogen-induced proliferation and growth of the functional endometrium occur primarily during the luteal phase and are dependent on the presence of functional PR expression. The absence of PR results in unopposed estrogen-induced endometrial hyperplasia in Pr knockout (PRKO) mice (Lydon et al. 1995). Tissue recombination studies with WT and PRKO uteri demonstrate that progesterone inhibits epithelial proliferation only in co-cultures with uteri expressing stromal PR (Kurita et al. 1998). Such studies support the importance of stromal PR expression as the inhibitory mediator of antiproliferative actions of progesterone. However, PR expression within the epithelia is still relevant as progesterone is unable to inhibit estradiol-induced endometrial proliferation or induce expression of important target genes encoding paracrine factors or cell cycle regulatory proteins in mice uteri lacking epithelial-specific PR expression (Franco et al. 2012). Therefore, the interplay between the epithelial and stromal cells of the endometrium is essential, with both cell types playing a role in the actions of progesterone.

Similar to the breast (discussed earlier in this review), each PR isoform can have very distinct target genes and biological functions, dependent on hormonal milieu and cellular context. In general, PR-B is considered as the stronger transcriptional activator and PR-A functions as a transcriptional inhibitor of PR-B activity (Tora et al. 1988, Vegeito et al. 1993, Hovland et al. 1998). Selective ablation of PR-A in mice results in a PR-B-dependent gain of function, with enhanced estradiol-induced endometrial proliferation (Mulac-Jericevic et al. 2000, Conneely et al. 2003). This unexpected observation suggests that PR-A is probably necessary for opposing the actions of both estradiol and progesterone in the endometrium, thereby limiting the proliferative effects of the PR-B receptor in this tissue. In addition, PR-A is also needed for progesterone-mediated changes during the luteal phase and implantation of the conceptus, as lack of PR-A results in impaired uterine implantation and little decidualization of the endometrial layer. This delicate balance of PR isoforms is further illustrated with transgenic mice overexpressing PR-A in glandular epithelium and stromal tissue. This experimental increase in the PR-A:PR-B ratio results in endometrial hyperplasia and atypia with enhanced expression of uterine epithelial growth factors such as amphiregulin known to be regulated by progesterone; these effects can be abolished by treatment with the antiprogestin, mifepristone (Fleisch et al. 2009). These results indicate that progesterone can be either an anti- or pro-proliferative force on the endometrium depending on isoform expression.

Studies on the uterine myometrium highlight the fact that the ratio of PR isoforms may be naturally exploited to remove the inhibitory effects of progesterone on myometrial contractions, thus allowing for estrogen activation and the initiation of parturition. One mechanism for this functional progesterone withdrawal may be a shift in the PR-A:PR-B ratio expressed within the myometrium with a concomitant antagonism of PR-B-mediated transcription (Pieber et al. 2001, Mesiano et al. 2002, 2011, Merlino et al. 2007). There is also evidence that a change in the PR-A:PR-B:PR-C isoform expression, specifically within the fundal myometrium (i.e., upper portion of the uterine body), could contribute to this process. Protein and mRNA expression of PR-C, as well as PR-B, increase during labor in women and are associated with NFκB activation and cytokine-mediated transcriptional activation of the PR gene (Condon et al. 2006). The potential transcriptional consequences of such isoform shifts, as experimentally manipulated or observed in these studies, are evident in gene array studies with primary human stromal cells expressing exogenous PR-A, PR-B, or the combination, where distinct expression profiles are observed for each isoform as well as progesterone concentration-dependent efficacy that was both target gene and isoform specific (Yudt et al. 2006). Overall, these results indicate that progesterone can be a positive or negative driver of cell processes such as endometrial proliferation or myometrial contractions depending on the isoform expression and downstream transcriptional and signaling activation. Misregulation of isoform expression, therefore, can lead to dysfunction and pre-neoplastic events.
Epithelial–stromal interactions within the endometrium: PR-driven paracrine communication

The regulation of paracrine factors and their signaling pathways by progesterone supports epithelial–stromal communication, which is critical for normal uterine function and may play a role in endometrial cancer pathogenesis (Fig. 3). Signaling via factors such as Indian hedgehog (Ihh) and Wnt ligands can be modulated by progesterone through regulation of the expression or activity of these paracrine factors or their downstream signaling molecules (Wetendorf & DeMayo 2012; Fig. 3). Within the hedgehog pathway, Ihh and dickkopf-related protein 1 (Dkk1) are PR target genes (Takamoto et al. 2002). Activation of stromal PR results in induction of Ihh expression by the epithelia and the subsequent stromal expression of patched homolog 1 (Ptc1) and nuclear receptor subfamily 2, group F, member 2 (Nr2f2) (Fig. 3). This can lead, in particular through NR2F2 (e.g., COUP-TFI), to activation of transcription factors such as Hand2 and potential antagonism of mitogenic pathway activation by growth factors, such as fibroblast growth factors (FGFs) (Li et al. 2011). This paracrine loop is thought to inhibit estrogen signaling and thereby halt uterine epithelial proliferation. The Wnt/β-catenin signaling pathway is critical for the control of stem cell/progenitor compartments and the balance between ‘stemness’ (e.g., proliferation with Wnt pathway active) and differentiation (e.g., inhibited Wnt pathway) in many tissues (Clevers 2006). In the endometrium, this pathway is also implicated in control of the proliferation–differentiation shift during the menstrual cycle and the actions of progesterone during the luteal phase may be through the inhibition of this pathway (Wang et al. 2010). Exposure to estrogen during the proliferative phase of the cycle leads to activation of this pathway with enhanced expression of Wnt pathway components (i.e., Wnt4, Wnt5a, Fzd2 (Hou et al. 2004)) and Wnt target genes such as IGF1 (Wang et al. 2009), a critical endometrial growth factor secreted by stromal cells (Cooke et al. 1997, McCampbell et al. 2006), as well as downregulation of DKK1 (stromal) and FOXO1, Wnt/β-catenin signaling inhibitors (Talbi et al. 2006, Wang et al. 2009; Fig. 3). Notably, Wnt4 is a paracrine effector for progesterone-induced expansion of the mammary stem cells (Joshi et al. 2010). Crosstalk with the PI3K/Akt pathway is also involved as E2-induced Akt activation, via ERz, results in inhibition of glycolgen synthase kinase (GSK-3β) and stabilization of β-catenin with enhanced transcription of Wnt target genes, ultimately leading to cell cycle progression (Tong & Pollard 1999).

Progesterone antagonizes the Wnt/β-catenin pathway via enhanced transcription of DKK1 and FOXO1 genes, retention of active GSK-3β, and nuclear exclusion of cyclin D1 resulting in cell cycle arrest (Chen et al. 2005, Ward et al. 2008, Wang et al. 2009, Kyo et al. 2011). Interestingly, blocking FOXO1 expression attenuates the ability of progesterone to inhibit epithelial cell growth, whereas expression of a dominant negative AKT enhances the inhibitory effect of this hormone (Kyo et al. 2011). These studies emphasize the crosstalk between paracrine signaling and mitogenic pathways modulated by ER and PR in the homeostasis of endometrial growth. Notably, the dysregulation of the PI3K/Akt and Wnt/β-catenin, in particular, is one hallmark of endometrial cancer pathogenesis. It is tempting to speculate that early events such as activating mutations in these key signaling pathways lead to imbalanced hormone-dependent stromal and epithelial crosstalk that then predisposes to neoplastic transformation of endometrial tissue.

Mechanisms of progestin resistance in endometrial cancer

Misregulation of PR isoform expression, localization, and activity are common phenotypes observed in EC that could be involved and potentially targeted to improve sensitivity to progestin therapy. In general, hyperplasias express higher levels of PR-A and PR-B (Miyamoto et al. 2004) and comparison of low- to high-grade endometrial cancers reveals reduced to absent expression of one or both isoforms in epithelia or stroma; these expression profiles are often associated with shorter progression-free survival and overall survival rates (Leslie et al. 1997, Miyamoto et al. 2004, Sakaguchi et al. 2004, Shabani et al. 2007, Jongen et al. 2009, Kreizman-Shefer et al. 2014). This silencing of PR expression may be due to hypermethylation of CpG islands within the promoter or first exon regions of the PR gene or due to the presence of associated deacetylated histones. These epigenetic modifications were observed in endometrial cancer cell lines as well as tumor samples and may be exclusive to PR-B (Sasaki et al. 2001, Xiong et al. 2005, Ren et al. 2007). Treatment of such cells with DNA methyltransferase or histone deacetylase inhibitors can restore both PR-B expression and its regulation of target genes such as FOXO1, p21 (CDKN1A), p27 (CDKN1B), and cyclin D1 (CCND1) (Xiong et al. 2005, Yang et al. 2014). Downregulation of PR via post-transcriptional mechanisms such as miRs could be another means of suppressing progesterone sensitivity, as observed in breast cancer cell
lines via overexpression of miR-26a and miR-181a (Maillot et al. 2009), but this remains to be examined in endometrial cancer models.

Post-translational modifications of PR, such as phosphorylation or SUMOylation, serve as input points for activated mitogenic pathways to regulate PR signaling (Dressing et al. 2009, Hagan et al. 2011b) and, therefore, may contribute to progesterone resistance. Studies with endometrial stromal cells have demonstrated that activation of cAMP signaling can sensitize cells to progesterone by suppressing SUMOylation of the PR-A isoform leading to enhanced transcriptional activity and target gene induction, supporting normal endometrial decidualization (Jones et al. 2006). Although the relevance of such PR modifications has not been extensively explored in the context of endometrial cancer, it is known that oncogenic activation of KRAS, PI3K, or AKT and/or loss of functional tumor suppressors such as PTEN are common genetic alterations observed in endometrial cancer (Hecht & Mutter 2006; Fig. 1). Janzen et al. (2013) have recently used an in vivo endometrial regeneration model to test how these common genetic alterations affect PR isoform expression and responsiveness to progesterone within epithelial and stromal compartments of the endometrium. Tumors generated from epithelial cells lacking PTEN were responsive to progesterone showing early decreased proliferation and later apoptosis, but co-administration of estrogen was necessary for tumor resolution as well as maintenance of stromal PR expression. Deletion of PR in stromal cells or combined epithelial-specific genetic mutations (i.e., PTEN loss and Kras activation) caused progesterone resistance, while overexpression of PR in stroma was able to re-sensitize tumors to therapy. Interestingly, tumors with the combined mutations showed depressed PR expression, especially stromal PR-A, due to epigenetic modifications; analysis of the PR-A promoter revealed multiple sites of hypermethylation. In addition to the function of stromal PR-A, studies have also highlighted the importance of PR-B where DNA methylation and decreased PR-B expression in endometrial cancer result in decreased FOXO1 and BIRC3 expression, enhancement of adhesion molecules, and cell cycle regulatory proteins. This ultimately lifts progesterone antagonism of estrogenic effects resulting in enhanced cell proliferation and survival (Shao 2013). These studies illustrate the importance of functional PR expression, uterine epithelial/tumor–stromal interactions, and hormonal milieu on PR signaling and therapeutic efficacy.

**Ovary**

**Epidemiological role of progesterone in ovarian tumors**

Ovarian cancer is the seventh most common cause of cancer-related deaths worldwide (Jemal et al. 2011). As the deadliest of all gynecologic malignancies, ovarian cancer has a death rate of more than 50% due to late detection and diagnosis of the disease and intrinsic or acquired resistance to current therapeutic regimens. The identification of robust biomarkers for early detection will have a substantial impact on survival rates, while prognostic molecular markers may allow for efficacious targeted therapeutic strategies.

A considerable body of epidemiological data suggests that progesterone and progestins play a protective role against ovarian carcinogenesis. Progesterone deficiencies due to increasing age, infertility, or a genetic LOH at the PR gene locus are associated with an increased risk of ovarian cancer (Gabra et al. 1996, Edmondson & Monaghan 2001). In contrast, elevated progesterone levels decrease the risk of ovarian cancer. The protective effect of pregnancy has been documented in Asian, European, and North American populations (Banks et al. 1997); progesterone levels during pregnancy are tenfold greater than luteal phase levels measured during the menstrual cycle. Similarly, hormonal oral contraceptive use has been consistently associated with a reduced risk. In an analysis of 20 epidemiological studies between 1970 and 1991, it was estimated that a 35% reduction in the risk was associated with ever-use of oral contraceptives (Hankinson et al. 1992). Additionally, the risk of ovarian cancer is correlated with the duration of oral contraceptive use: 10–12% decrease in the risk with 1 year of use and 50% decrease after 5 years of use in both nulliparous and parous women (Hankinson et al. 1992). Progesterone exerts a protective effect on the risk of ovarian cancer by reducing ovulation through elevated progesterone levels from oral contraceptive use or during pregnancy. Furthermore, PR expression, PR-B specifically (Akahira et al. 2000, 2002, Lenhardt et al. 2012), in ovarian tumors is a favorable prognostic marker associated with longer progression-free survival (Hempling et al. 1998, Akahira et al. 2000, Munstedt et al. 2000, Lindgren et al. 2001, Lee et al. 2005, Hodgall et al. 2007, Tangjitgamol et al. 2009, Yang et al. 2009, Sinn et al. 2011).

BRCA1/2 mutations may alter the production and sensitivity to estrogen and progesterone as carriers have an increased risk for breast and ovarian cancer. Studies on mice carrying a Brca1 mutation in ovarian granulosa
A recent study has demonstrated that ovulation, the release of hormones (e.g., estrogen and progesterone), growth factors, and inflammatory factors among others, promoted the migration of intrauterine-injected malignant cells toward the ovarian stromal compartment to form ‘ovarian’ tumors (Yang-Hartwich et al. 2014). Thus, it is plausible that the unique hormonal milieu provided by functional ovaries serves to attract pre-malignant and malignant cells that may remain dormant (i.e., under progesterone concentrations) or fully progress to tumors (i.e., post-menopausal contexts or upon loss of progesterone or functional PRs). Approximately 90% of ovarian cancers are detected in the ovary, with over 50% ovarian cancers diagnosed in post-menopausal women (American Cancer Society, 2014. Cancer Facts and Figures 2014. Atlanta).

Until recently, little has been known about the relative distribution of PR within the subtypes of epithelial ovarian tumors. In a cohort of 504 tumors, we reported that 35% of ovarian tumors are PR positive, with the highest total PR expression in endometrioid (67%) and serous (35%; low-grade serous, 64%) subtypes (Diep et al. 2013). In accordance with our study, the international Ovarian Tumor Tissue Analysis consortium examined the association of ER and PR expression with subtype-specific survival in ~3000 invasive epithelial ovarian tumors reporting positive total PR expression in endometrioid (67%), low-grade serous (57%), and high-grade serous (31%) tumors (Sieh et al. 2013). Additionally, the study confirmed the prognostic significance of PR expression in ovarian tumors strongly expressing PR (≥50% tumor cell nuclei staining). Strong PR expression in high-grade serous ovarian carcinomas was associated with a significant improvement in survival; positive PR expression (weak or strong) in endometrioid carcinomas was associated with significantly improved disease-specific survival independent of patient age and tumor grade, site, and stage. Notably, ER expression conferred a patient survival advantage in endometrioid ovarian tumors only. ER may contribute to the favorable prognosis in endometrioid ovarian tumors via regulation of PR expression; a functional ER signaling pathway promotes robust PR expression. While total PR levels are routinely measured in breast and endometrial cancers (but rarely in ovarian cancer) for clinical management and disease treatment, very few studies have examined the levels of PR isoforms in ovarian tumors. To our knowledge, only three studies (Akahira et al. 2000, 2002, Lenhard et al. 2012) have reported differential expression of PR isoforms in ovarian tumors. These studies have reported a dominance of PR-B expression in ovarian tumors across all sub-types, with PR-B frequently expressed in the serous subtype. In contrast, PR-A expression was weakly expressed in mucinous and serous ovarian carcinomas and comparison of normal ovarian tissues with malignant ovarian tissues.
revealed reduced to absent expression in malignant tumors relative to PR-B (Akahira et al. 2002).

**Progesterone actions in ovarian cancer**

The molecular mechanisms of progesterone’s protective role in ovarian cancer are not well understood; both proliferative and inhibitory actions of progesterone have been reported in ovarian cancer cell line models. Several independent *in vitro* studies demonstrated antiproliferative actions of progesterone at higher concentrations (≥ 1 μM) in ovarian cancer cells, primarily through the induction of apoptosis (Bu et al. 1997, Keith Bechtel & Bonavida 2001, Yu et al. 2001, Syed & Ho 2003), while fewer studies reported progesterone as proliferative in these cells at lower concentrations (Syed et al. 2001, Fauvet et al. 2006). The opposing cellular responses of ovarian cancer cells to progesterone may be attributed to cell context-dependent regulatory inputs to PR (discussed earlier in this review), such as progesterone dosing, kinase activation state of the cells, cofactor availability, or PR-A and PR-B ratios. Ovarian cancer cells are susceptible to concentration-dependent and biphasic effects within the same cell model systems as mentioned previously for uterine and breast (in 2D culture systems) cancer cells. Similar to breast and uterus, crosstalk between PR and growth factor-mediated signaling pathways (i.e., protein kinases) presumably directs PR promoter selection and specific cell fates (e.g., apoptosis). The relative abundance of cofactors that associate with PR also varies in a tissue-specific manner (Giangrande et al. 2000, Han et al. 2005). As in other tissues (discussed earlier in this review), shifts in PR isoform ratios (PR-A and PR-B) and cofactor availability may contribute to variations in biological responses to progesterone.
PR isoform-specific actions are largely undefined in ovarian cancer. However, our recent study has defined a mechanism for PR-B regulation of ovarian cancer cellular senescence in response to progesterone. Using ovarian cancer cell models, we demonstrated that ligand-activated PR-B acting through a FOXO1-dependent mechanism induced p21, a known mediator of cellular senescence. FOXO1, a transcriptional factor, has been demonstrated to interact physically with other nuclear steroid hormone receptor proteins, such as AR (Li et al. 2003, Fan et al. 2007), ER-α (Schuur et al. 2001), and both PR isoforms (Kim et al. 2005, Rudd et al. 2007). Our study demonstrated that PR-B and FOXO1 were co-recruited to a PRE-containing region in the upstream promoter of p21 upon progesterin (R5020) treatment. Both proteins were required to cooperatively activate progesterin-induced p21 expression and induce PR-dependent cellular senescence. PR-B appears to be a more potent driver of ovarian cancer cell senescence relative to PR-A; PR-B but not PR-A induces robust FOXO1 expression (Diep CH, Knutson TP, and Lange CA, unpublished observations).

As stated above for breast studies, we suspect that PR isoforms in ovarian cancer models are also exquisitely sensitive to kinase inputs that may alter this biological outcome. Both PR-B and FOXO1 are tightly regulated by phosphorylation events. Hormone-driven breast and gynecological cancers frequently exhibit upregulated protein kinases, such as MAPK (Fairve et al. 2005), CDK2 (Pierson-Mullany & Lange 2004), and CK2 (Hagan et al. 2011a), which directly phosphorylate and modulate PR-B target gene selectivity (Fig. 1). Notably, the same kinases that are recruited to PR-B in ‘rapid’ signaling (i.e., extranuclear) complexes (i.e., CDK2 and MAPK) also inhibit FOXO1 via regulation of specific phosphorylation sites that favor nuclear export (Hedrick et al. 2012). Dysregulation of FOXO1 is associated with tumorigenesis and cancer progression. FOXO1 is downregulated in several carcinomas, including ovarian cancer (Goto et al. 2008), through alterations in upstream regulators, post-translational dysregulation, or by genetic mutations (Myatt & Lam 2007). Specifically, AKT-mediated serine/threonine phospho-regulation of FOXO1 is well defined and prevents FOXO1 nuclear accumulation, thus impairing target gene regulation (Myatt & Lam 2007). As mutations of PI3Ks or PTEN are common early events in cancer (particularly in breast, uterine, and ovarian cancers), activated AKT and other mitogenic protein kinases may prevent PR-induced senescence signaling by nuclear exclusion of FOXO1. Thus, the early loss or inactivation of FOXO1 may render PR ‘incompetent’ at genes required for the induction of cellular senescence, leading to the loss of protective ‘sensing’ by progesterone in ovarian tumors. Whether these events may redirect PR to ‘alternate’ genes that instead favor tumor progression is unknown and a topic for further study.

Finally, mortality rates for ovarian cancer have remained largely unaffected despite clinical advances in detection methods, surgical techniques, and treatment regimens. Although extensive surgery followed by chemotherapy is often effective at inducing clinical remission, the treatment is toxic and rarely results in a cure. Other treatment regimens, such as hormonal therapy, have been evaluated for ovarian cancer. The use of progestins alone (megestrol acetate and medroxyprogesterone acetate) as ovarian cancer therapies has been examined in several relatively small phase II clinical trials with variable inclusion criteria and modest response rates (Modugno et al. 2012). However, retrospective studies evaluating the association of total PR expression and progression-free disease survival (Hempling et al. 1998, Akahira et al. 2000, Munstedt et al. 2000, Lindgren et al. 2001, Lee et al. 2005, Hogdall et al. 2007, Tangjitgamol et al. 2009, Yang et al. 2009, Sinn et al. 2011, Sieh et al. 2013) support the concept that subsets of PR-positive ovarian tumors are highly sensitive to hormones and thus more likely to respond to endocrine therapy.

Overall, identifying the mechanisms governing PR-A vs PR-B-specific gene regulation may provide insight for exploiting the protective actions of progesterone in PR-positive gynecological tumors to induce growth arrest and ultimately favor cell death, namely the development of PR isoform-specific ligands may allow for promotion of PR-B-driven cellular senescence in ovarian cancer or induction of the protective actions of PR-A in uterine cancer. Growth-arrested senescent cells cannot further divide, but depend upon specific kinase-mediated signal transduction pathways for prolonged survival, and thus may be more vulnerable to subsequent therapies that inhibit mitogenic protein kinases and thereby promote apoptosis. Thus, as part of novel combination therapies, PR-targeted strategies could provide a safe and useful means to improve treatment outcomes and increase overall patient survival.

Antiprogestins in preclinical and clinical development

Table 1 depicts antiprogestins currently under preclinical and clinical development in breast cancer, endometrial cancer, endometriosis, leiomyomas, and ovarian cancer. Mifepristone (RU486) has been studied in several phases I
and II clinical trials for breast and gynecological diseases and cancers as it blocks the transcriptional activity of PR by directly binding to and recruiting corepressors to PR (depending on cellular context) (Han et al. 2007).

Paradoxically, mifepristone was originally developed as a potent antiglucocorticoid compound and was later discovered to have antiprogesterone activity when mifepristone caused termination of pregnancy in preclinical studies (Spitz & Bardin 1993). Similarly, mifepristone can also bind to the AR (Song et al. 2004). While the structures of progesterone, glucocorticoid, and ARs are very similar, the varying affinity of mifepristone to these steroid receptors may account for the limited efficacy and substantial toxicity observed in several clinical trials for breast and ovarian cancer (Perrault et al. 1996, Rocereto et al. 2010). A new generation of PR antagonists attenuates malignant proliferation of tumors and is highly selective for PR with potent antiprogesterone activity but minimal antiglucocorticoid effects in vitro and in vivo studies. These PR antagonists include APR19, CDB-2914 (ulipristal), CDB-4124 (telapristone), J867 (asoprisnil), ORG31710, WAY-255348, ZK230211 (lonaprisan), ZK98299 (onapristone), and a 17-fluorinated steroid branded as EC304 (Table 1) (reviewed in Chabbert-Buffet et al. 2005, Spitz (2006), Knutson & Lange (2014) and Goyeneche & Telleria (2015)). Ultimately, the development of highly selective PR antagonists, and the identification of patient cohorts that will benefit from antiprogestins and their use in combination with other endocrine therapies may significantly advance hormone-modulation strategies for breast and gynecological cancers.

**Summary of discussion**

Herein, we have discussed the pivotal role of altered progesterone signaling in the development and progression of hormone-regulated tumors. In the breast, progesterone promotes a proliferative and pro-survival response (i.e., PR is a major downstream effector of estrogen signaling), but inhibits estrogen-induced growth in the reproductive tract. The paradoxical effects of progesterone observed in tumors arising from these tissues may be largely dependent on endogenous cell context and the tissue

---

### Table 1  Current antiprogestins in preclinical and clinical development in breast and gynecological diseases

<table>
<thead>
<tr>
<th>Antiprogestin</th>
<th>Phase</th>
<th>Disease</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>APR19</td>
<td>Preclinical</td>
<td>Breast cancer</td>
<td>Khan et al. (2013)</td>
</tr>
<tr>
<td>EC304</td>
<td>Preclinical</td>
<td>Breast cancer</td>
<td>Nickisch et al. (2013)</td>
</tr>
<tr>
<td>ORG31710</td>
<td>Preclinical</td>
<td>Breast cancer</td>
<td>Bakker et al. (1990)</td>
</tr>
<tr>
<td>WAY-255348</td>
<td>Preclinical</td>
<td>Breast cancer</td>
<td>Yudt et al. (2011)</td>
</tr>
<tr>
<td>Asoprisnil (J867)</td>
<td>II</td>
<td>Endometriosis</td>
<td>DeManno et al. (2003) and Chwalisz et al. (2007)</td>
</tr>
<tr>
<td>Lonaprisan (BAY86-5044, ZK230211)</td>
<td>II</td>
<td>Breast cancer</td>
<td>Jonat et al. (2013)</td>
</tr>
<tr>
<td>Mifepristone (RU486)</td>
<td>I-II</td>
<td>Breast cancer</td>
<td>Romieu et al. (1987), Klijn et al. (1989) and Perrault et al. (1996)</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>Endometrial cancer</td>
<td>Ramondetta et al. (2009)</td>
</tr>
<tr>
<td></td>
<td>II</td>
<td>Leiomyoma</td>
<td>Engman et al. (2009) and Yerushalmi et al. (2014)</td>
</tr>
<tr>
<td>Onapristone (ZK98299)</td>
<td>II</td>
<td>Ovarian cancer</td>
<td>Rocereto et al. (2000) and Rocereto et al. (2010)</td>
</tr>
<tr>
<td></td>
<td>I</td>
<td>Breast cancer</td>
<td>Helle et al. (1998) and Robertson et al. (1999)</td>
</tr>
<tr>
<td>Telapristone (CDB-4124, Proellex)</td>
<td>II</td>
<td>PR+ tumors</td>
<td>Gupta et al. (2013)</td>
</tr>
<tr>
<td>Ulipristal (CDB-2914)</td>
<td>II-III</td>
<td>Breast cancer</td>
<td>Ioffe et al. (2009)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Endometriosis</td>
<td>Jonat et al. (2013)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Leiomyoma</td>
<td>Levens et al. (2008)</td>
</tr>
</tbody>
</table>

### Table 2  Summary of PR isoform actions

<table>
<thead>
<tr>
<th>Tissue type</th>
<th>Isoforms</th>
<th>Isoform-specific actions of progesterone</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast</td>
<td>PR-A</td>
<td>Trans-represses PR, ER, AR, and GR activities</td>
</tr>
<tr>
<td></td>
<td>PR-B</td>
<td>Required for normal mammary gland development and expansion</td>
</tr>
<tr>
<td>Uterus</td>
<td>PR-A</td>
<td>Required for normal uterine development and function</td>
</tr>
<tr>
<td></td>
<td>PR-B</td>
<td>Dominant isoform in normal stromal cells</td>
</tr>
<tr>
<td>Ovary</td>
<td>PR-A</td>
<td>Essential for normal ovarian function</td>
</tr>
<tr>
<td></td>
<td>PR-B</td>
<td>Dominant isoform in ovarian carcinomas</td>
</tr>
</tbody>
</table>

---
microenvironment, namely the opposing effects of progesterone may be attributed to altered expression or activity of PR isoforms, the contextual interactions between the epithelial and stromal compartments observed in breast and endometrial tissues, changes in their relative regulation either by post-translational modifications or via differential crosstalk with cofactor-binding partners that serve as major inputs to altered transcriptional activity and promoter selection in the various target tissues (see Table 2 for an overview).

Although elegant models have recently emerged (Karst et al. 2011, Tanos et al. 2013), knowledge gaps still exist. What are the best methods and experimental models to elucidate progesterone-specific effects in hormone-responsive tumors? While breast, endometrial, and ovarian cancers are diagnosed in both pre- and post-menopausal populations, a majority of the current cell-based models were originally established from post-menopausal patients. To understand steroid receptor actions, cells are treated with varying concentrations of exogenous hormones that may or may not reflect true physiological levels experienced in a pre-menopausal (cyclical hormone exposure) or post-menopausal (constant/low hormone exposure) context. Are the hormone concentrations used in the laboratory relevant to these contexts and thus to the biology of the tumors that arise? In addition, decreased PR expression is associated with progression of disease in breast and gynecologic cancers (Gross et al. 1984, Balleine et al. 1999), whereas over 50% of acquired endocrine-resistant breast tumors retain PR expression (Encarnacion et al. 1993, Johnston et al. 1995). How do breast and other tumors lose PR expression and/or regain it during extended periods of endocrine (antiestrogen therapy)? How should we model these changes? Concerning in vitro models, PR expression is often lost when primary isolates or immortalized cell lines are continuously cultured on 2D surfaces. The development of co-culture or 3D models may more accurately reflect in vivo cellular architecture relevant to paracrine signaling and tumor biology (Lo et al. 2012) and will allow a more accurate characterization of the mechanisms and biological effects of hormone and antitumor treatments. Finally, routine detection and quantification of individual PR isoforms in clinical samples may provide valuable information as potentially distinct biomarkers of tumor behavior that could be used to further guide endocrine therapy.

Understanding how PRs function differentially in each normal and neoplastic tissue type will reveal how these highly modified receptors can be therapeutically targeted, perhaps as separate isoforms, to favor one biological outcome (growth inhibition, senescence, and apoptosis) over another (proliferation and survival). Ultimately, in order to effectively manipulate PR action pharmacologically to treat tumors arising from different tissue types, we must first appreciate their mechanistic complexity. Isoform-specific ligands as activators or inhibitors would be a valuable set of tools to accomplish this goal. In the current age of cancer genomics and personalized medicine, clinical readouts of PR-driven gene signatures may provide an additional means to discern context-dependent protective vs deleterious PR actions present in individual tissues and tumors.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

Funding
This work was supported by NIH grant R01 CA159712 (to C A L), a supplement to the parent NIH grant R01 CA15972-51 (to C A L), Cancer Biology Training Grant NIH T32 CA09138 (to C H D) and National Center for Advancing Translational Sciences of the National Institutes of Health Award ULTR000114 (to C H D).

References


Charles D 1964 Iatrogenic endometrial patterns. *Clinical Pathology* 17 205–212. (doi:10.1136/cp.17.3.205)


Daniel AR, Gaviglio AL, Knutson TP, Ostrander JH, D’Assoro AB, Ravindranathan P, Peng Y, Raj GV, Yee D & Lange CA 2015...


Hou X, Tan Y, Li M, Dey SK & Das SK 2004 Canonical Wnt signaling is critical to estrogen-mediated uterine growth. Molecular Endocrinology 18 3035–3049. (doi:10.1210/me.2004-0259)


Received in final form 15 December 2014
Accepted 12 January 2015
Accepted Preprint published online 13 January 2015